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**PFAS Analysis Based Upon a pH-Variable LC  
Mobile Phase Gradient**

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Babcock Laboratories, Inc.



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# Outline

- **Background**
  - CA Investigative Orders
  - Distinct Analyte List (new analytes)
  - Methods Gap
- **Analytical Scalability**
  - Current analyte chemistry and technique
- **Expanded Utility of pH Gradient**



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# CA Investigative Orders

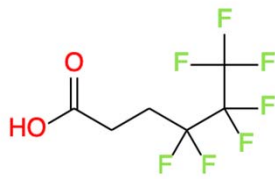
Recent Timeline

- March 20, 2019 - Landfills and Airports
  - 38 analytes with RLs at 5 – 8ng/L. (23 “required”)
  - New analytes not in any validated method from EPA or ASTM (C16, C18, FOSE, x:3-FTCA)
- October 25, 2019 – Chrome Platers
  - Same 38 analytes. (25 “required”)
  - Nov 22, 2019 – Extension granted
- July 9, 2020 – POTW
  - 4 short chain analytes added (533 add-ons: PFEESA, NFDHA, PFMBA, PFMPA)
- March 12, 2021 – Bulk Fuel Storage / Refineries
  - 31 analytes. (31 required)
  - Removed x:3-FTCA, PFHxDA, PFOcDA, 10:2-FTS, PFNS

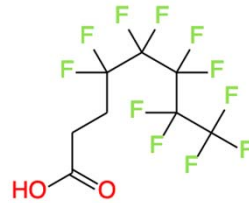


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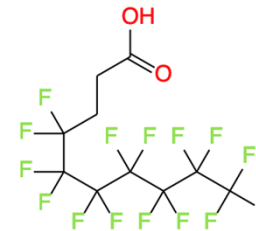
## Newer PFAS Chemistry Not In Current Methods



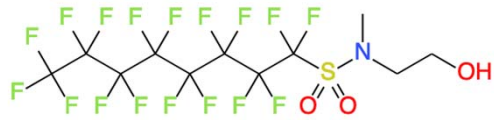
3:3-FTCA



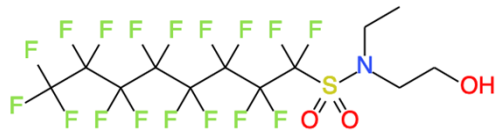
5:3-FTCA



7:3-FTCA



N-MeFOSE  
(non-ionic)



N-EtFOSE  
(non-ionic)



N-MeFOSA  
(pKa~3.47)



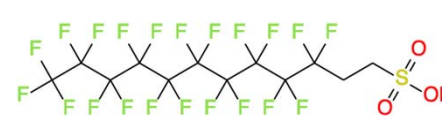
N-EtFOSA  
(pKa~3.64)



PFHxDA (C16)



PFOcDA (C18)

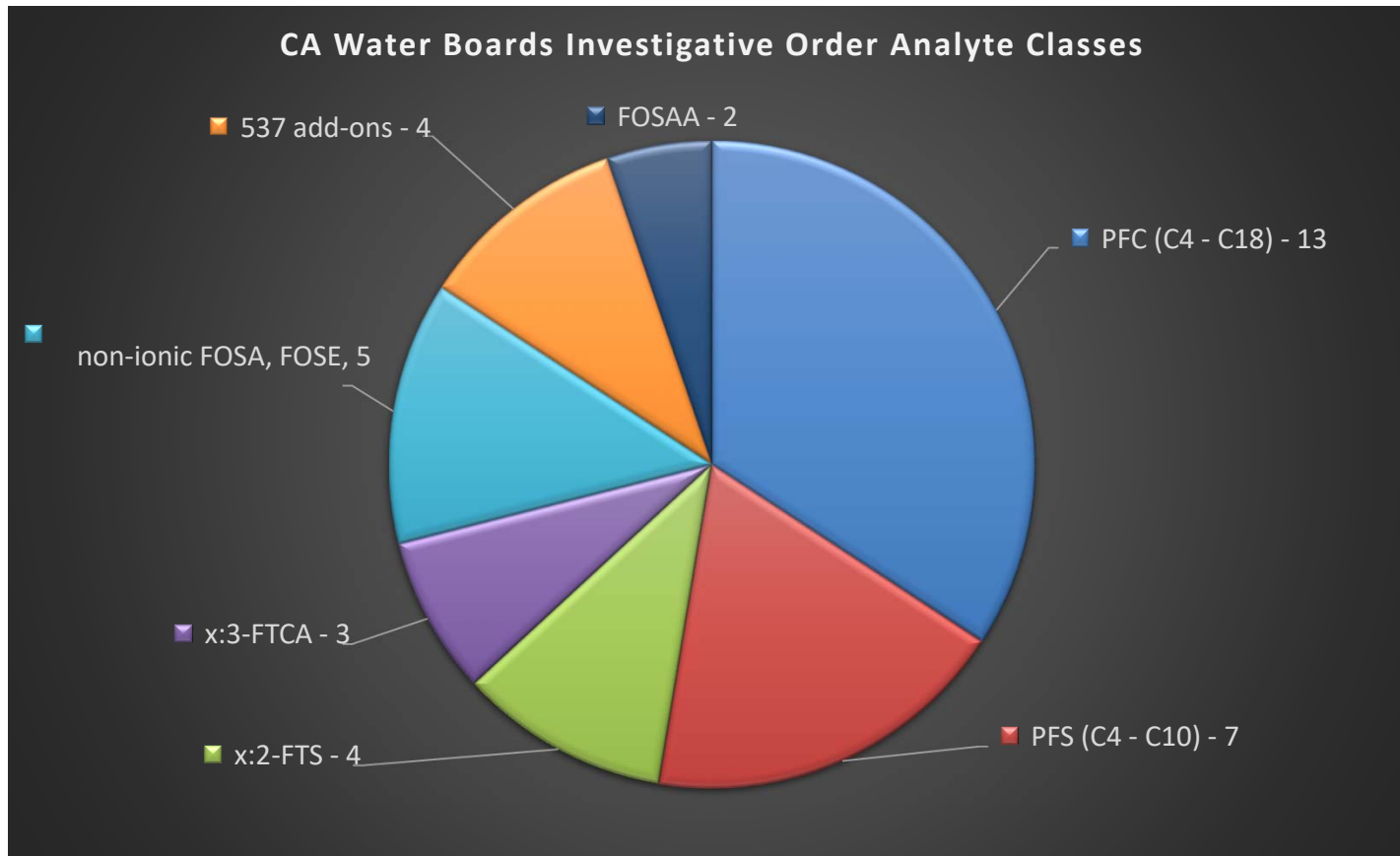


10:2-FTS



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# Analyte Chemistry





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# Which Method?

- **Non-Potable Water and PFAS**
  - No standard EPA methods for PFAs in wastewater existed and the first (EPA 8327) was not promulgated yet
  - 537- Mod not standardized / “Mod” not within method scope
  - ASTM methods
    - May have scalability issues with long chain PFAS (D7979)
    - Can’t be used to comply with QSM 5.3 for trace level PFAS
  - Dept of Defense (DoD) released their QSM 5.3 (2019)
    - Has become a gold standard in the United States
    - Requires Solid Phase Extraction (SPE) and cleanup (eg: GCB)
    - Can Scale analyte List
    - **Became the choice of the CA Water Boards**



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## Method Chemistry

| Method     | % Organic | Perfluorocarboxylate Range |
|------------|-----------|----------------------------|
| EPA 537.1  | 96% MeOH  | C6 to C14                  |
| EPA 533    | 80% MeOH  | C4 to C12                  |
| EPA 8327   | 50% MeOH  | C4 to C14                  |
| ASTM D7979 | 50% MeOH  | C4 to C14                  |

- EPA 533 and 537.1 prescribe extract composition
- Ease of use for EPA 8327 / ASTM D7979 is that it's a 1:1 dilution (50% Organic)
- QSM 5.3 does not require a particular % Organic in the extract although SPE and cleanup is required



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# Scalability/Constraints

- Need to do PFBA and PFOcDA (C4 to C18)
  - PFOcDA has solubility issues in >10% water
    - Finding PFOcDA in water samples is unlikely
    - Extract must be 100% organic → can't use ASTM D7979
  - PFBA is difficult to chromatograph in 100% organic
    - Lower LC gradient starting pH to 3-4 to promote protonated form
  - Long chain PFAS stick to column at low pH
    - Increase pH (>9) during LC gradient to minimize tailing
  - Most LC columns not compatible with high pH
    - Use Phenomenex Kinetex EVO product
- All other PFAS fit well into this framework although non-ionic better suited for GC-MS/MS
  - In LC-MS, N-MeFOSE and N-EtFOSE detected as  $[M+HOAc-H]^-$
  - Product ion is acetate at mass 59 (not as selective)





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# Data Quality Objectives

- Meet DOD QSM 5.3 + Table B-15
  - Calibration/ICV/CCVs (standard)
  - Daily instrument sens. check at LOQ +/-50%
  - Retention Times
    - RT vs Exp RT (+/- 0.4min)
    - RT quant ion vs qual ion (+/- 2sec)
    - RT vs isotope RT (eg: PFOA and 13C-PFOA +/- 0.1min)
  - Ion Ratios 50-150% expected ratios
  - Use specific transitions for specific analytes unless matrix is demonstrated and documented

RT Stability  
Required

\*\*\*Not an exhaustive list of all requirements



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# Instrument Method /Extraction

**Table 1 - MS Conditions**

|            |                    |             |
|------------|--------------------|-------------|
| Mass Spec  | Thermo TSQ Vantage |             |
| Cap Temp   | 250                | C           |
| Vap Temp   | 300                | C           |
| Sheath gas | 40                 | arb         |
| Aux Gas    | 15                 | arb         |
| Sweep Gas  | 0                  | arb         |
| CID        | 1.0                | mTorr Argon |
| Q1 FWHM    | 0.4                | Da          |

**Table 2 - HPLC Conditions**

HPLC Agilent 1100  
Column C18 EVO 100x2.1mm  
Delay Column C18 EVO 50x2.1mm ★  
Column Temp 35C  
Inj Vol 10uL  
★ Eluent A 20 mM Acetic Acid  
★ Eluent B 25 mM Ammonium Hydroxide in Methanol

| Time  | % A | % B |
|-------|-----|-----|
| 0.00  | 95  | 5   |
| 1.20  | 55  | 45  |
| 3.60  | 35  | 65  |
| 11.00 | 10  | 90  |
| 13.00 | 10  | 90  |
| 13.01 | 95  | 5   |
| 17.00 | 95  | 5   |

**Table 3 - SPE Sample Prep**

Cartridge Strata-WAX/GCB 200/50mg

Sample Check pH. Buffer using phosphate if not between 6-7

Condition 10mL 0.1% NH<sub>4</sub>OH in MeOH  
10mL MeOH  
10mL Phosphate Buffer pH=7

Load 250mL

Wash 5mL 0.1% Formic Acid in 50:50 Water: Methanol

Dry 2 mins

Elute 4mL 0.1% NH<sub>4</sub>OH in MeOH

Soak 2 mins

Elute 4mL 0.1% NH<sub>4</sub>OH in MeOH

Vap Using N<sub>2</sub> just below 2mL

Adjust to 2mL using 100% MeOH

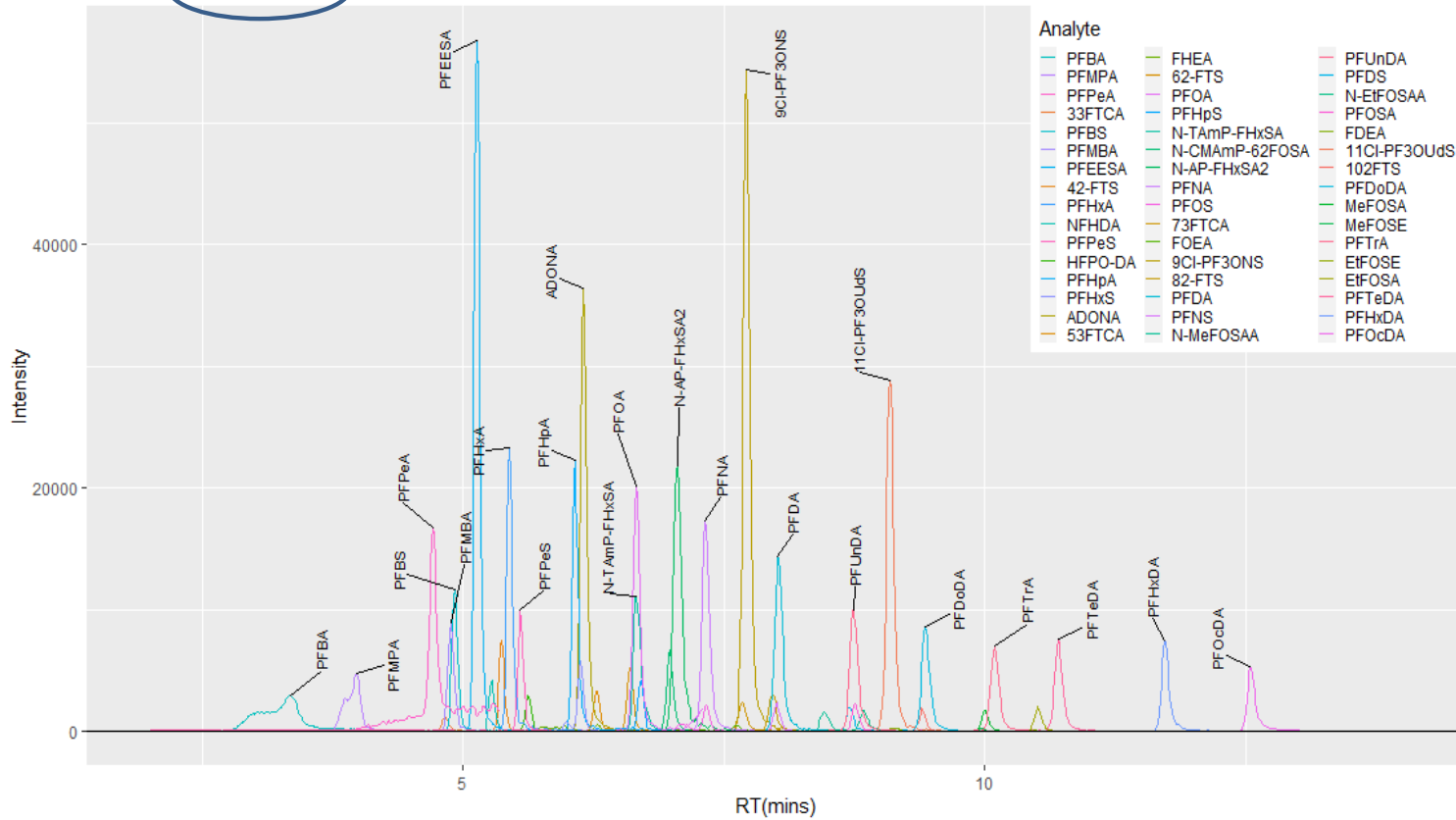


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# 20mM Ammonium Acetate

## Total Ion Chromatogram

5ppb Std in 100% MeOH

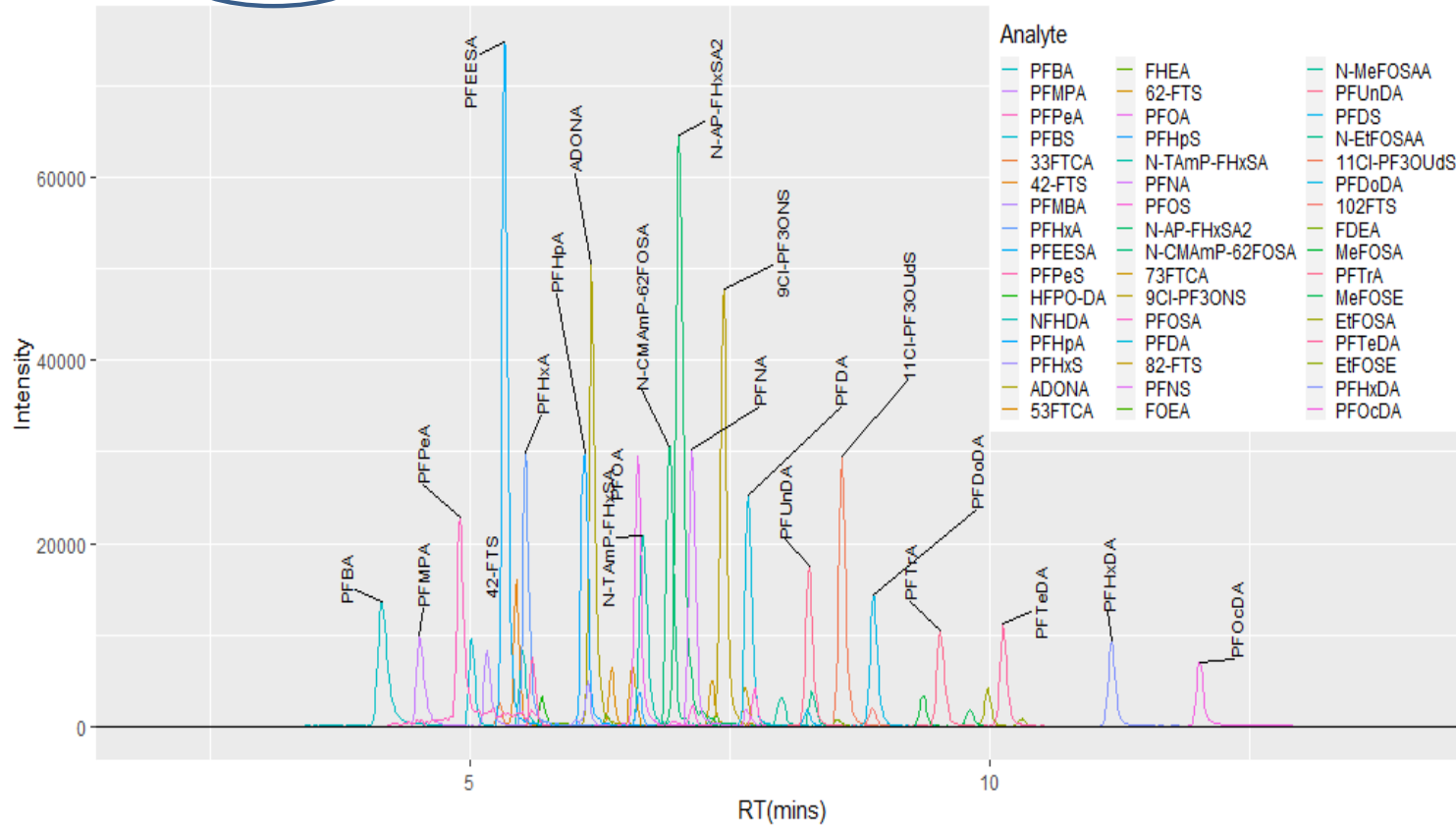




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# pH Gradient Elution

Total Ion Chromatogram  
5ppb Std in 100% MeOH





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## Details of pH Gradient

| 20mM HOAc in Water | 25mM NH <sub>4</sub> OH in MeOH |           |
|--------------------|---------------------------------|-----------|
| %A                 | % B                             | Actual pH |
| 100%               | 0%                              | 3.6       |
| 95%                | 5%                              | 3.9       |
| 90%                | 10%                             | 4.2       |
| 80%                | 20%                             | 4.6       |
| 70%                | 30%                             | 5.1       |
| 60%                | 40%                             | 5.8       |
| 50%                | 50%                             | 6.4       |
| 40%                | 60%                             | 7.1       |
| 35%                | 65%                             | 8.2       |
| 30%                | 70%                             | 8.5       |
| 20%                | 80%                             | 9.0       |
| 10%                | 90%                             | 9.3       |
| 0                  | 99.5%                           | >10       |

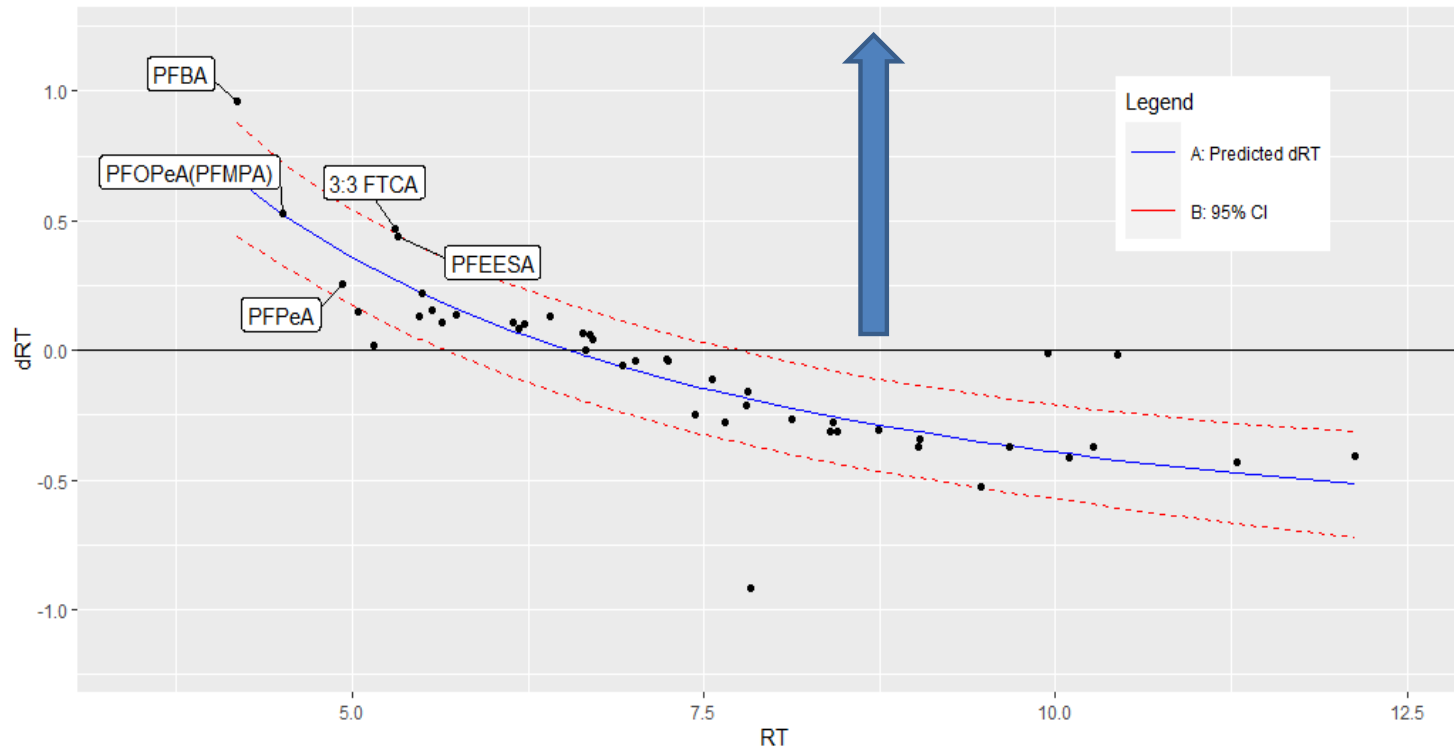
- Ammonium Acetate dissolved in water ***not*** a buffer
- Actual pH during run subject to delay volume
- Still provides stable RTs (meets QSM 5.3)



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# Retention Increase – $\Delta RT > 0.25$

Delta-RT of pH Gradient vs. 20mM NH<sub>4</sub>OAc (pH=7)  
5ppb Std in 100% MeOH

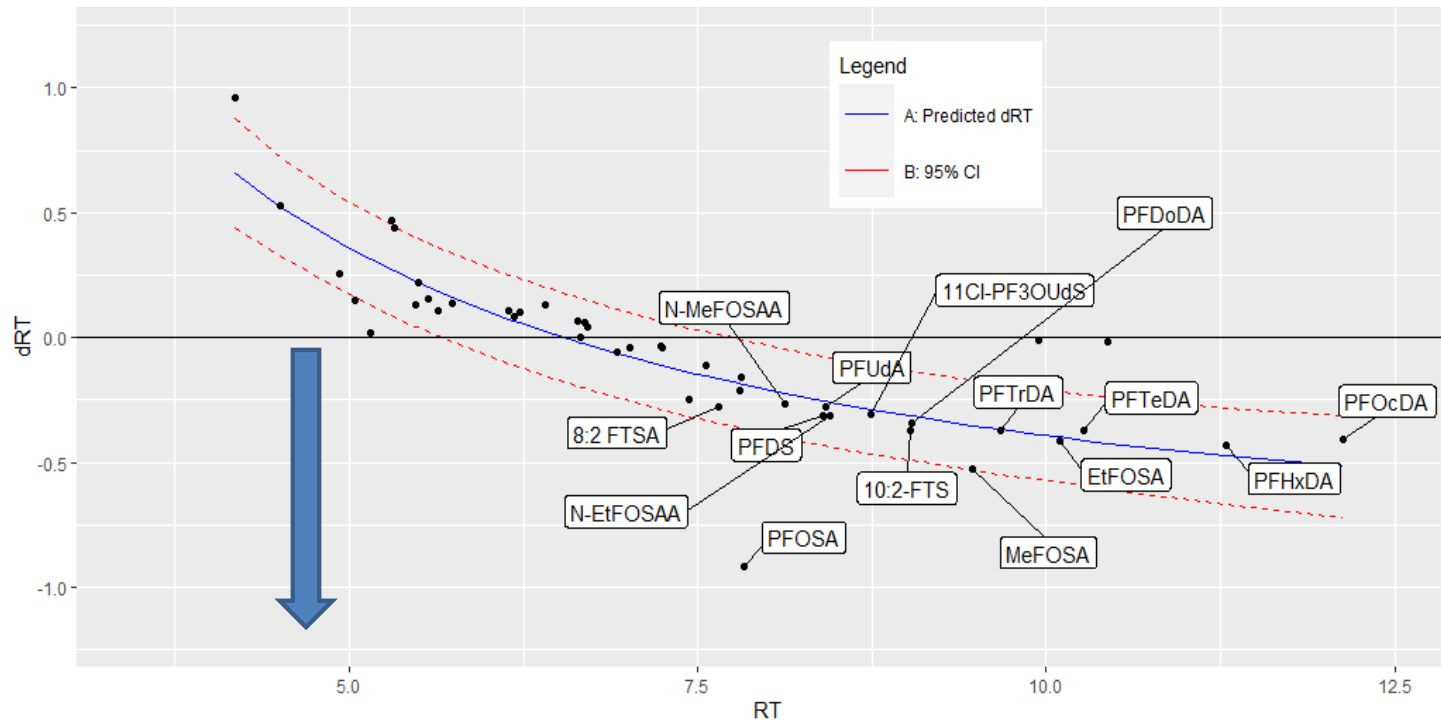




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# Retention Decrease – $\Delta RT < 0.25$

Delta-RT of pH Gradient vs. 20mM NH<sub>4</sub>OAc (pH=7)  
5ppb Std in 100% MeOH

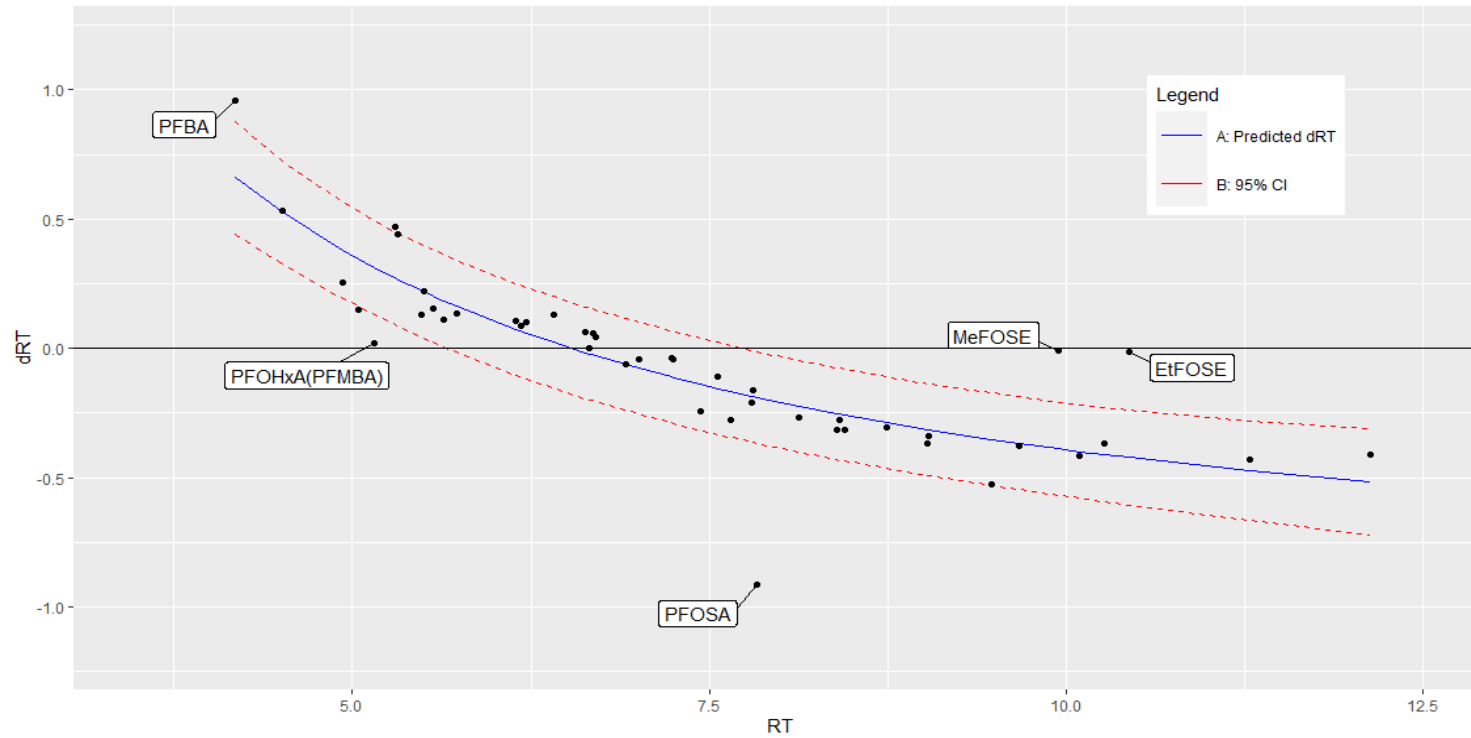




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# Significant Selectivity Changes Using GVLMA Method

Delta-RT of pH Gradient vs. 20mM NH<sub>4</sub>OAc (pH=7)  
5ppb Std in 100% MeOH

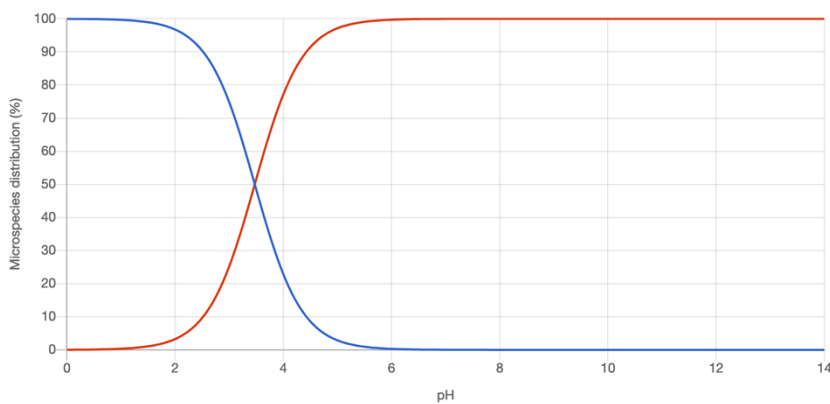




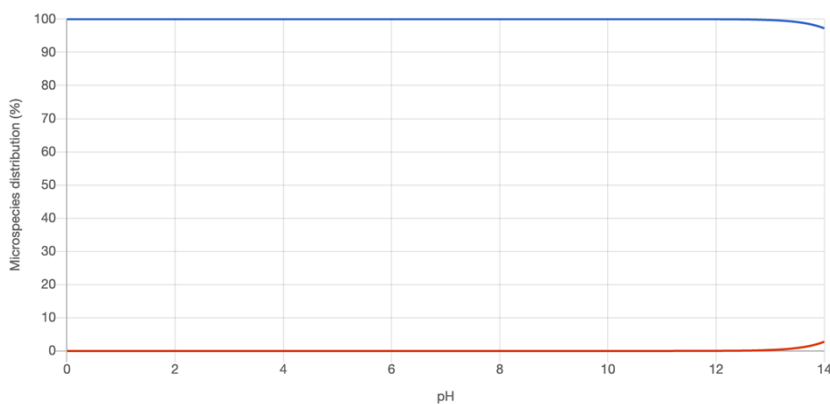


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## N-MeFOSA vs. N-MeFOSE Ionization and pH



N-MeFOSA  
(pKa~3.47)



N-MeFOSE  
(non-ionic)

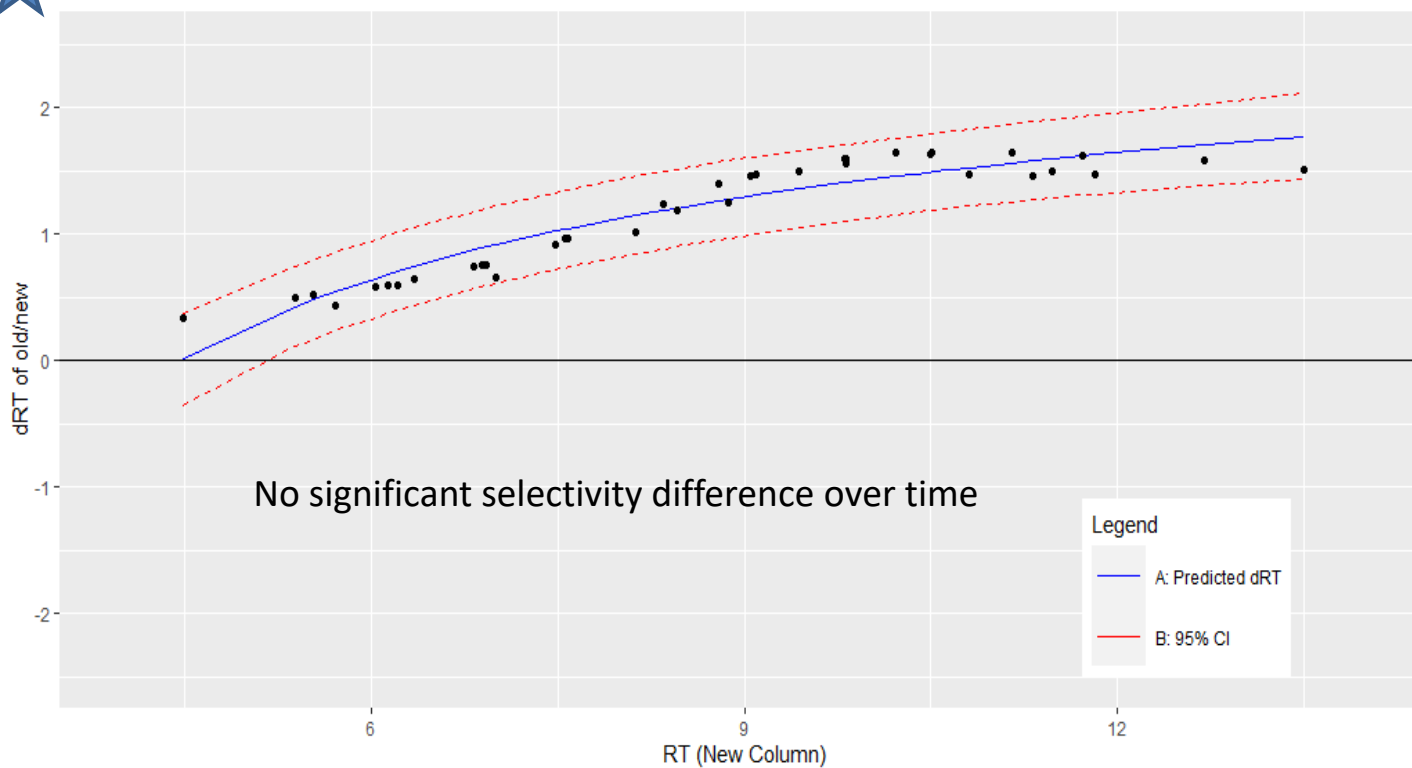
Data from Chemicalize.com



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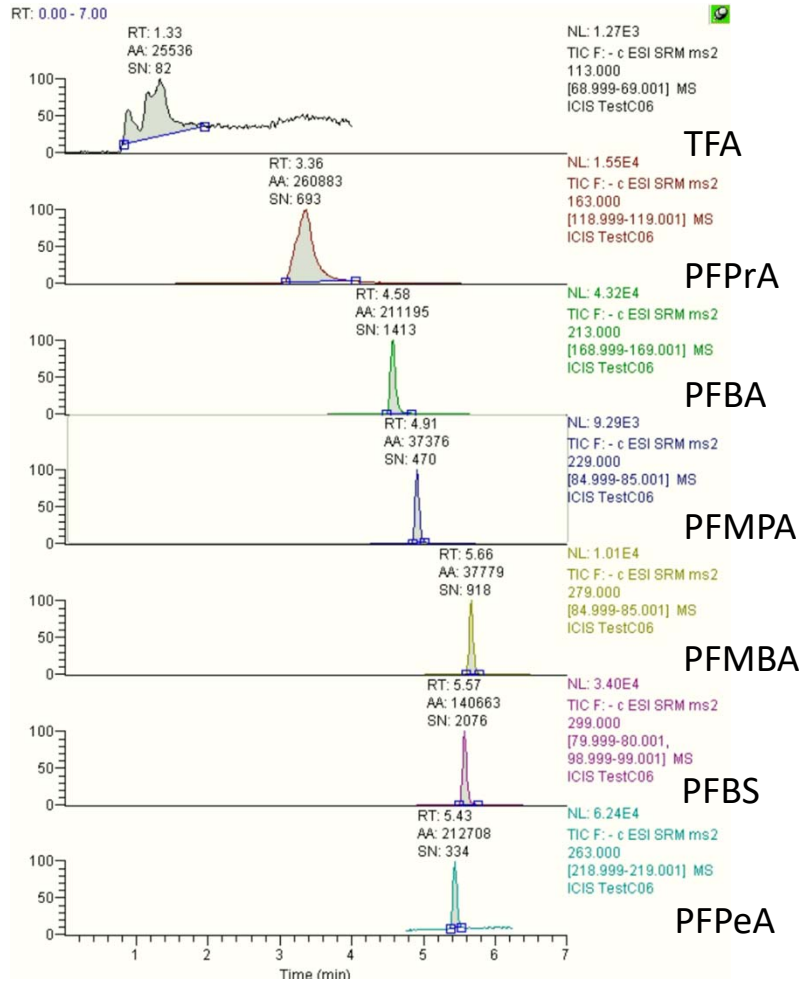
# New/Old Column – RT Stability

★ Delta RT Plot using the Varied-pH Gradient  
Kinetex C18 EVO - New vs 6mo used





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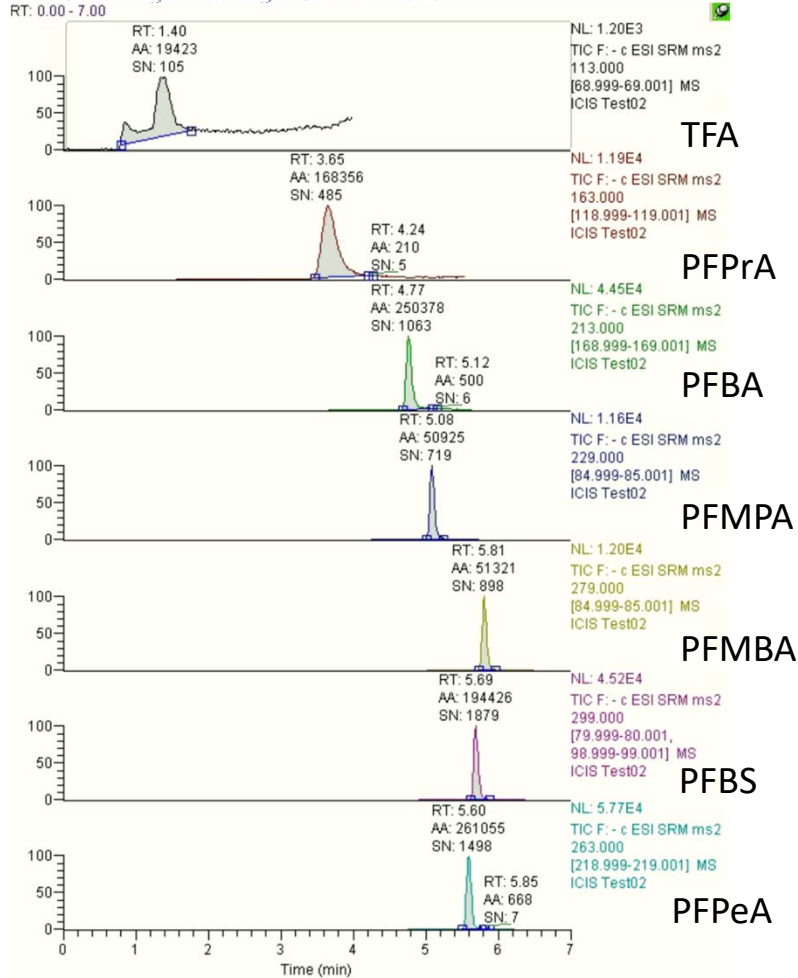


## Short Chain PFAS (< C6)

- Ultra-Short short chain PFAS retention and sensitivity unacceptable for TFA (C2)
- TFA was lost in standards due to volatility and may have analytical precision issues



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# What about Formic Acid (20mM HOFo)

Lower pKa of Formic acid provides increased retention of short chain however, MeFOSE and EtFOSE not as sensitive because they are detected as adducts of Formate



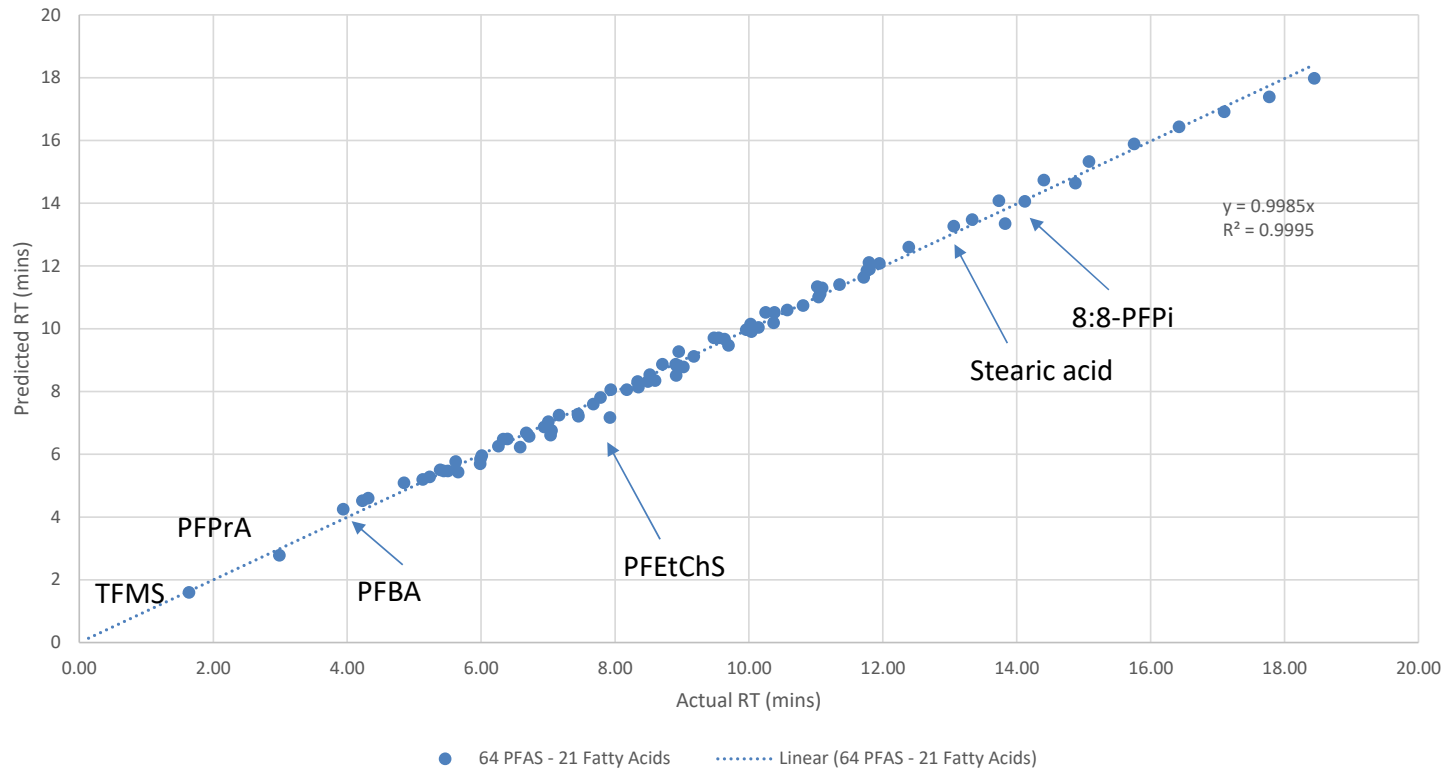
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# Useful for RT Prediction

Proprietary QSRR Model

*“All models are wrong,  
but some are useful”*

Target-Class PFAS RT Model  
Actual vs. Predicted





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## Utility of a pH Gradient

- Increased Retention of Short Chain PFAS
- Was validated under QSM 5.3 (see other work)
- Better Retention Time Prediction of Short Chain PFAS
  - More accurate models based on computationally predicted values (e.g. SLogP)
  - Help Qualitative ID in Targeted Screening using High-Res MS
    - Must know structure to compute SLogP



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## More Technical Info



fluoroalkyl  
(PFAS) Extraction by



PFAS Analysis Based Upon a  
pH-Variable LC Mobile Phase  
Gradient

- GVLMA Statistical method to determine selectivity difference
- Release Full Method Validation soon

[phenomenex.com/pfas](https://www.phenomenex.com/pfas)



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# Acknowledgements

## Contributors/Reviewers

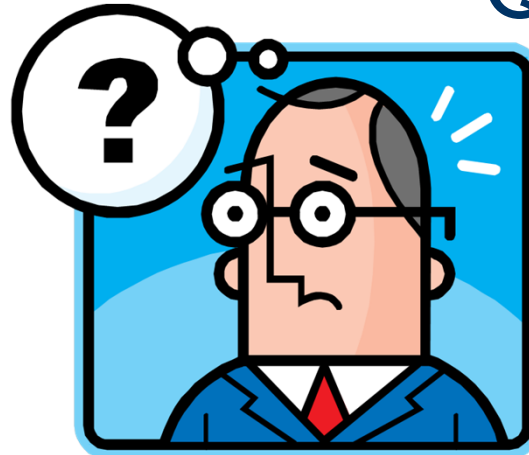
- Richard Jack - Phenomenex
- Sam Lodge - Phenomenex
- David Kennedy - Phenomenex





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# Questions?



Contact:

|               |  |
|---------------|--|
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**Thank You!**